

**In the Specification**

Please replace the paragraph beginning on page 19, line 14 with the following amended paragraph:

Based on these data and excluding the six cysteines, we judge that the KuDom structure will allow those substitutions shown in Table 9. The class indicates whether the substitutions: A) are very likely to give a stable protein having substantially the same binding to hNE, hCG, or some other serine protease as the parental sequence, B) are likely to give similar binding as the parent, or C) are likely to give a ~~protein~~protein retaining the KuDom structure, but which are likely to affect the binding. Mutants in class C must be tested for affinity, which is relatively easy using a display-phage system, such as the one set forth in W0/02809. The affinity of hNE and hCG inhibitors is most sensitive to substitutions at positions 15, 16, 17, 18, 34, 39, 19, 13, 11, 20, 36 of BPTI, if the inhibitor is a mutant of ITI-D1, these positions must be converted to their ITI-D1 equivalents by aligning the cysteines in BPTI and ITI-D1.

Please replace the paragraph beginning on page 35, line 2 with the following amended paragraph:

**Example 9: Amino-acid sequences of EPI-HNE-3 and EPI-HNE-4**

Table 10 gives amino acid sequences of four human-neutrophil-elastase (hNE) inhibitor proteins: EPI-HNE-1 (which is identical to EpiNE1), EPI-HNE-2, EPI-HNE-3, and EPI-HNE-4. These proteins have been derived from the parental Kunitz-type domains shown. Each of the proteins is shown aligned to the parental domain using the six cysteine residues (shaded) characteristic of the Kunitz-type domain. Residues within the inhibitor proteins that differ from those in the parental protein are in upper case. Entire proteins having the sequences EPI-HNE-1, EPI-HNE-2, EPI-HNE-3, and EPI-HNE-4 (Table 10) have been produced. Larger proteins that comprise one of the hNE-inhibiting sequences are expected to have potent hNE-inhibitory activity; EPI-HNE-1, EPI-HNE-2, EPI-HNE-3, and EPI-HNE-4 are particularly preferred. It is expected that proteins that comprise a significant part of one of the hNE-inhibiting sequences found in Table 10 (particularly if the sequence starting at or before the first cysteine and continuing through or beyond the last cysteine is retained) will exhibit potent hNE-inhibitory activity.

Please replace the paragraph beginning on page 36, line 11 with the following amended paragraph:

We have used this high density *Pichia pastoris* production system to produce proteins by secretion into the cell ~~CM~~culture medium. Expression plasmids were constructed by ligating synthetic DNA sequences encoding the *S. cerevisiae* mating factor  $\alpha$  prepro peptide fused directly to the amino terminus of the desired hNE inhibitor into the plasmid pHIL-D2 using the *Bst*BI and the *Eco*RI sites shown. Table 24 gives the DNA sequence of a *Bst*BI-to-*Eco*RI insert that converts pHIL-D2 into pHIL-D2(MF $\alpha$ -PrePro::EPI-HNE-3). In this construction, the fusion protein is placed under control of the upstream inducible *P. pastoris aox1* gene promoter and the downstream *aox1* gene transcription termination and polyadenylation sequences. Expression plasmids were linearized by *Sac*I digestion and the linear DNA was incorporated by homologous recombination into the genome of the *P. pastoris* strain GS115 by spheroplast transformation. Regenerated spheroplasts were selected for growth in the absence of added histidine, replated, and individual isolates were screened for methanol utilization phenotype (*mut* $^+$ ), secretion levels, and gene dose (estimated via Southern hybridization experiments). High level secretion stains were selected for production of hNE inhibitors: PEY-33 for production of EPI-HNE-2 and PEY-43 for production of EPI-HNE-3. In both of these strains, we estimate that four copies of the expression plasmid are integrated as a tandem array into the *aox1* gene locus.

Please replace Table 21, page 89 with the following amended Table:

TABLE 21

ITI-D1-derived hNE Inhibitors										
WEAK ( $K_D > 10^{-8}$ M)										
1...5....0....5....0....5....0....5....0....5....0....5...										
1.	KED	SCQLG	YSAGPC	MGMT	SRYF	YNGT	SMAC	ETFQY	GGCMGN	NNFVTEKDCLQTCRGA
MODERATE ( $10^{-8} > RD 22 10^{-9}$ )										
2.	KED	SCQLG	YSAGPC	<b>VAMF</b> PRYF	YNGT	SMAC	ETFQY	GGCMGN	NNFVTEKDCLQTCRGA	
3.	<b>RPDF</b> CQLG	YSAGPC	<b>VAMF</b> PRYF	YNGT	SMAC	ETFQY	GGCMGN	NNFVTEKDCLQTCRGA		
STRONG ( $10^{-9} > KD > 10^{-11}$ D)										
4.	<b>RPDF</b> CQLG	YSAGPC	<b>VAMF</b> PRYF	YNGT	SMAC	ETFQY	GGCMGN	NNFVTEKDCLQTCRGA		
5.	<b>RPDF</b> CQLG	YSTG	PC	<b>VAMF</b> PRYF	YNGT	SMAC	ETFQY	GGCMGN	NNFVTEKDCLQTCRGA	
6.	KED	FCQLG	YSAGPC	<b>VAMF</b> PRYF	YNGT	SMAC	ETFQY	GGCMGN	NNFVTEKDCLQTCRGA	
7.	KED	SCQLG	YSAGPC	<b>VAMF</b> PRYF	YNGT	SMAC	ETFQY	GGCMGN	NNFVTEKDCLQTCRGA	
8.	<b>RPDF</b> CQLG	YSAGPC	<b>IGMF</b> SR	YNGT	SMAC	ETFQY	GGCMGN	NNFVTEKDCLQTCRGA		
VERY STRONG ( $K_D < 10^{-11}$ M)										
9.	<b>RPDF</b> CQLG	YSAGPC	<b>VAMF</b> PRYF	YNGT	SMAC	OTF	<b>VY</b> GGCMGN	NNFVTEKDCLQTCRGA		
10.	<b>RPDF</b> CQLG	YSAGPC	<b>VAMF</b> PRYF	YNGA	SMAC	OTF	<b>VY</b> GGCMGN	NNFVTEKDCLQTCRGA		
11.	<b>RPDF</b> CQLG	YSAGPC	<b>VAMF</b> PRYF	YNGT	SMAC	ETF	<b>VY</b> GGCMGN	NNFVTEKDCLQTCRGA		
12.	<b>RPDF</b> CQLG	YSAGPC	<b>VGMF</b> SR	YNGT	SMAC	OTF	<b>VY</b> GGCMGN	NNFVTEKDCLQTCRGA		

Residues shown underlined and bold are changed from those present in ITID1

Sequences Key:

1. ITI-D1	SEQ ID NO. 008 <u>16</u>
2. ITI-D1E7	SEQ ID NO. 009 <u>21</u>
3. BITI	SEQ ID NO. 030 <u>141</u>
4. BITI-E7	SEQ ID NO. 010 <u>142</u>
5. BITI-E7-1222	SEQ ID NO. 012 <u>143</u>
6. AMINO1	SEQ ID NO. 015 <u>22</u>
7. AMINO2	SEQ ID NO. 016 <u>23</u>
8. MUTP1	SEQ ID NO. 014 <u>24</u>
9. BITI-E7-141	SEQ ID NO. 011 <u>17</u>
10. MUTT26A	SEQ ID NO. 018
11. MUTQE	SEQ ID NO. 017
12. MUT1619	SEQ ID NO. 013 <u>20</u>

Please replace Table 65, beginning on page 72 to page 73, with the following amended Table:

**Table 659: Definition of Class A, B and C mutations in PCT/US92/01501.**

Classes:      A      No major effect expected if molecular charge stays in range -1 to +1.  
                  B      Major effects not expected, but are more likely than in "A".  
                  C      Residue in the binding interface; any change must be tested.  
                  X      No substitution allowed.

Res.      EpiNE1

Id.	(SEQ ID NO:7)	Substitutions	Class
1	R	any	A
2	P	any	A
3	D	any	A
4	F	Y, W, L	B
5	C	C	X
6	L	non-proline	A
7	E	L, S, T, D, N, K, R	A
8	P	any	A
9	P	any	A
10	Y	non-proline pref'r'd	B
11	T	any	C
12	G	must be G	X
13	P	any	C
14	C	C strongly preferred, any non-proline	C
15	I	V, A	C
16	A		C
17	F	L, I, M, Y, W, H, V	C
18	F	Y, W, H	C
19	P	any	C
20	R	non-proline pref'r'd	C
21	Y	F & Y most pref'r'd; W, I, L pref'r'd; M, V allowed	C
22	F	Y & F most pref'r'd; non-proline pref'r'd	Y, F B
23	Y	Y & F strongly pref'r'd	F, Y B
24	N	non-proline pref'r'd	A
25	A	any	A

26	K	any	A
27	A	any	A
28	G	non-proline prefr'd	A
29	L	non-proline prefr'd	A
30	C	must be C	X
31	Q	non-proline prefr'd	B
32	T	non-proline prefr'd	B
33	F	F very strongly prefr'd; Y possible	X
34	V	any	C
35	Y	Y most prefr'd; W prefr'd; F allowed	B

Res.

Id.	EpiNE1	Substitutions	Class
36	G	G strongly prefr'd; S, A prefr'd;	C
37	G	must be G so long as 38 is C	X
38	C	C strongly prefr'd	X
39	M	any	C
40	G	A,S,N,D,T,P	C
41	N	K,Q,S,D,R,T,A,E	C
42	G	any	C
43	N	must be N	X
44	N	S,K,R,T,Q,D,E	B
45	F	Y	B
46	K	any non-proline	B
47	S	T, N, A, G	B
48	A	any	B
49	E	any	A
50	D	any	A
51	C	must be C	X
52	M	any	A
53	R	any	A
54	T	any	A
55	C	must be C	X
56	G	any	A
57	G	any	A
58	A	any	A

prefr'd stands for preferred.

Please replace Table 11, beginning on page 80 to page 81, with the following amended Table:

Table 11: Restriction sites in plasmid pHIL-D2

pHIL-D2, 93-01-02      Ngene = 8157

Non-cutters

AfI <sub>II</sub>	ApaI	AscI	AvaI	AvrII	BamHI	BglII
Bsp120I	BsrGI	BssHII	BstEII	FseI	MluI	NruI
PacI	PmlI	RsrII	SacII	SexAI	SfiI	SgfI
SnaBI	SpeI	Sse8387I	XhoI (PaeR7I)		XmaI (SmaI)	

Cutters

AatII GACGTc	1	5498
AfI <sub>III</sub> Acrygt	1	7746
AgeI Accggt	1	1009
BlpI GCtnagc	1	597
BspEI (BspMII, AccIII) Tccgga	1	3551
BspMI gcaggt	1	4140
Bst1107I GTAtac	1	7975
BstBI (AsuII) TTcgaa	2	945 4780
Bsu36I CCtnagg	1	1796
Ecl136I GAGctc	1	216
EcoRI Gaattc	1	956
EspI (Bpu1102I) GCtnagc	1	597
HpaI GTTaac	1	1845
NcoI Ccatgg	1	3339
NdeI CAtatg	1	7924
NsiI (Ppu10I) ATGCAt	1	684
PflMI CCANNNNntgg	1	196 (SEQ ID NO:127)

PmeI	GTTTaaac	1	420
PstI	CTGCAg	1	6175
PvuI	CGATcg	1	6049
SapI	gaagagc	1	7863
SacI	GAGCTc	1	216
SalI	Gtcgac	1	2885
ScaI	AGTact	1	5938
SphI	GCATGc	1	4436
StuI	AGGcct	1	2968
SwaI	ATTaaat	1	6532
Tth111I	GACNnnngtc	1	7999
XbaI	Tctaga	1	1741
XcmI	CCANNNNnnnntgg	1	711 (SEQ ID NO:128)

Aox1 5' 1 to about 950

Aox1 3' 950 to about 1250

His4 1700 to about 4200

Aox1 3' 4500 to 5400

bla 5600 to 6400

f1 ori 6500 to 6900

Please replace Table 630 on page 123 with the following amended Table:

**Table 63040:** Temperature stability of EPI-HNE proteins

Temperature (°C)	Residual hNE Inhibitory Activity			
	EPI-HNE-1	EPI-HNE-2	EPI-HNE-3	EPI-HNE-4
0	97	101	96	100
23	100	103	105	103
37	100	97	99	98
45	103			
52		101	100	
55	99			98
65	94	95	87	
69				82
75	100			
80		101	79	
85	106			63
93		88	57	
95	64			48

Proteins were incubated at the stated temperature for 18 hours in buffer at pH 7.0. In all cases protein concentrations were 1  $\mu$ M. At the end of the incubation period, aliquots of the reactions were diluted and residual hNE-inhibition activity determined.